

Antifungal Activity of Selenium Nanoparticles and Selenium Disulfide against Two *Malassezia* Species

Faranak Mavandadnejad¹, Fatemeh Rafii², Elnaz Faghfuri¹, Narges Mokhtari-Nori¹
Sasan Rezaie³, Ahmad Reza Shahverdi^{1*}

¹Department of Pharmaceutical Biotechnology and Biotechnology Research Center, Faculty of Pharmacy
Tehran University of Medical Sciences, Tehran, Iran

²Division of Microbiology, National Center for Toxicological Research, FDA, Jefferson, AR 72079, USA

³Department of Medical Mycology and Parasitology, School of Public Health, Tehran
University of Medical Sciences, Tehran, Iran

shahverd@tums.ac.ir

Abstract: Fungi in the genus *Malassezia* are involved in skin disorders, including dandruff and seborrheic dermatitis, two commonly recurring scalp conditions that are often treated with selenium-containing materials. The antifungal effects of SeS₂ and SeNPs against *M. sympodialis* and *M. furfur* were compared. The results showed that SeNPs had higher antifungal activity than SeS₂ against both species. Both preparations were more effective against *M. sympodialis* than against *M. furfur*. Post-antifungal effects were not observed for *M. furfur* after short exposure to either SeNPs or SeS₂. However, cultures treated for a short time with the MIC or lower than the MIC of SeNPs or SeS₂ showed better growth than untreated cultures following incubation with 1/10 MIC or less of either of the compounds. Brief exposure of *Malassezia* spp. to less than the MFC of SeNPs or SeS₂ may result in the proliferation of fungal cells that are less sensitive to selenium and may contribute to the recurrence of infection following treatment.

Keywords: Selenium nanoparticles, *Malassezia*, antifungal activity, post-antifungal effect

INTRODUCTION

Yeasts of the genus *Malassezia* are lipophilic fungi of the normal skin flora of humans and other warm-blooded animals, but they may also be the cause of skin disorders, including dandruff and seborrheic dermatitis [Aggarwal, et al., 2003; Anwar et al., 2016; Roques, et al., 2006; Rudramurthy et al., 2014]. Seborrheic dermatitis (SD), covering areas of the scalp, face and trunk, is a superficial inflammatory skin disorder, which affects 1%-10% of the population [Roques et al., 2006]. These diseases are treated with antimycotic shampoos containing various antifungal agents, including selenium disulfide (SeS₂), which has antifungal activity against a variety of fungi, including *Malassezia* spp. [Aggarwal, et al., 2003; Chu et al., 1984; McGinley et al., 1982; Van Cutsem et al., 1990]. A low concentration of selenium is essential to living organisms [Kitajima et al., 2013]. As a trace element, it is incorporated in the amino acids of selenoproteins and antioxidant enzymes and it protects cells from free radicals. However, in higher concentration, it inhibits fungal growth [Keiliszek et al., 2015; Kitajima et al., 2013]. The reason for the antifungal activity of selenium in *Malassezia* has not been studied but Wu et al [Wu et al., 2014] found that a high concentration of selenium damages the cellular oxygen-eliminating system of *Penicillium expansum*, which leads to an increase in the production of intracellular ROS. In *P. expansum*, the inhibitory effect is related to the selenium concentration used [Wu et al., 2014]. SeS₂ has been shown to have antifungal activity against *Malassezia* [Aggarwal, et al., 2003; McGinley et al., 1982]. However, after treatment with antifungal agents, including SeS₂, SD tends to relapse. Aggarwal et al treated 20 SD patients once a week for three weeks with 2.5% SeS₂ shampoo and one month after therapy found that three of the patients had mild or considerable residual disease. There is a need for finding more effective compounds for the treatment of S.D. and investigating the reason for the recurrence of infection after SeS₂ treatment.

Antifungal Activity of Selenium Nanoparticles and Selenium Disulfide Against Two *Malassezia* Species

In recent years, interest in the biological activities of metal nanoparticles has increased and the use of metal nanoparticles as antimicrobial agents has been investigated [Anwar et al., 2016; Ormland et al., 2004; Ren et al., 2009; Sadiq et al., 2009; Tan et al., 2009; Wang et al. 2005; Wang et al. 2007; Gijjar et al., 2009]. SeNPs have been reported to have antioxidant activity [Ormland et al., 2004; Tan et al., 2009; Wang et al. 2005; Zeng et al., 2008; Zhang et al., 2001]. We previously have shown that SeNPs biosynthesized by *Klebsiella pneumoniae* have antifungal activities against several fungi, including *Malassezia* spp [Shahverdi et al., 2010].

In this study, we compared the antifungal effect of SeS₂ and SeNPs for *M. furfur* and *M. sympodialis* and determined the MFC that results in eradication of these species. Since scalp SD is treated with a selenium-containing shampoo, we also investigated the post-antifungal effect of short exposures of *M. furfur* to SeS₂ and SeNPs on the subsequent growth of this fungus.

MATERIALS AND METHODS

Synthesis of SeNPs

Biogenic SeNPs were freshly prepared and purified using a method previously described [Shahverdi et al., 2010, Fesharaki et al., 2010]. Briefly, an inoculum of *Klebsiella pneumoniae* [Fesharaki et al., 2010] was prepared by transferring a single colony from a tryptic soy agar plate to TSB (Merck, Darmstadt, Germany) and growing the culture at 37°C to an OD₆₀₀ of 1.0. Fresh TSB, pH 7.2, supplemented with 200 mg/l Se⁴⁺ (equal to 559.19 mg of selenium chloride) was inoculated with 1% (v/v) of a *K. pneumoniae* culture and incubated at 37°C for 24 h. *K. pneumoniae* cells containing red selenium particles were disrupted by autoclaving for 20 min at 121°C and 1.2 kg/cm² pressure. The released SeNPs were centrifuged at 25,000 x g for 15 min and washed three times with distilled water. The washed sample and SeS₂ were sonicated for 10 min (Tecna6, Tecno-Gaz Industries, Parma, Italy). The sizes of the SeNPs and SeS₂ used during this study were less than 250 nm, as determined by TEM [Shahverdi et al., 2010, Fesharaki et al., 2010].

Comparison of antifungal activity of SeNPs and SeS₂ for *Malassezia* spp.

Clinical strains of *M. sympodialis* and *M. furfur* were obtained from the Culture Collection of the Department of Medical Mycology and Parasitology, School of Public Health, Tehran University of Medical Sciences, Tehran (Iran). The effects of biogenic SeNPs and SeS₂ (Sigma-Aldrich, Germany) on the survival of *M. sympodialis* and *M. furfur* were separately tested in LNB supplemented with different concentrations (0-300 µg/ml) of SeNPs or SeS₂ according to the antifungal susceptibility testing recommendations of the CLSI. The broth cultures were inoculated with 10⁶ CFU/ml of each fungal species and incubated for 120 h at 30°C. The viability of cells of each strain remaining in each culture was determined by plating the culture on LNA and counting the resulting colonies.

MIC and MFC Determination

A conventional serial dilution method was used to determine MIC of both selenium preparations for *M. sympodialis* and *M. furfur* in LNB according to the CLSI. The media contained either SeNPs or SeS₂ in a concentration range of 0 to 300 µg/ml, and were inoculated with approximately 10⁶ CFU/ml of each species. The minimum concentrations of SeNPs and SeS₂ capable of inhibiting visible growth of the fungal strains during 96-120 h of incubation at 35°C were determined and reported as MICs. The MFC of SeNPs and SeS₂ was measured by plating the cultures used for MIC determinations onto selenium-free LNA plates. After incubation of plates at 35°C for 96-120 h, the lowest concentration of SeNPs or SeS₂ that killed 99.0-99.5% of the cells and resulted either in no growth or fewer than three colonies on the plates, was considered the MFC [Espinol et al., 2002].

Antifungal Activity of Selenium Nanoparticles and Selenium Disulfide Against Two Malassezia Species

Post-antifungal effect (PAFE) of SeNPs and SeS₂

The effect of pre-exposure of *M. furfur* to (50µg/ml) SeNPs and SeS₂ on its growth after removal of the drug was determined by a modification of the Odenholt-Tornqvist method [Espinol et al., 2002]. as follows: SDB medium (Merck,Darmstadt, Germany) containing 50µg/ml of SeNPs and SeS₂ was inoculated with 5 × 10⁶ cells/ ml of *M. furfur* separately and incubated for 2 h at 35°C. For controls, cultures of *M. furfur* were grown in SDB medium without selenium. The cultures were diluted with pre-warmed SDB at a ratio of 1/10 and incubated at 35°C for 120 h. Then, the viability of *M. furfur* in cultures with and without selenium compounds was determined by plating the samples on LNA medium and counting the CFUs. All samples were homogenized before CFU determination. The experiments were repeated three times.

RESULTS AND DISCUSSION

In this study, we compared the *in vitro* antifungal activity and post-antifungal effect of SeNPs and SeS₂. We found higher antifungal potency of SeNPs for *M. furfur* and *M. sympodialis* in comparison with SeS₂ and showed that neither of the compounds had a PAFE for *M. furfur*. To compare the antifungal activities of biogenic SeNPs and SeS₂, the viability of *M. sympodialis* and *M. furfur* after incubation with different concentrations of these compounds was determined (Figure 1, Figure 2).

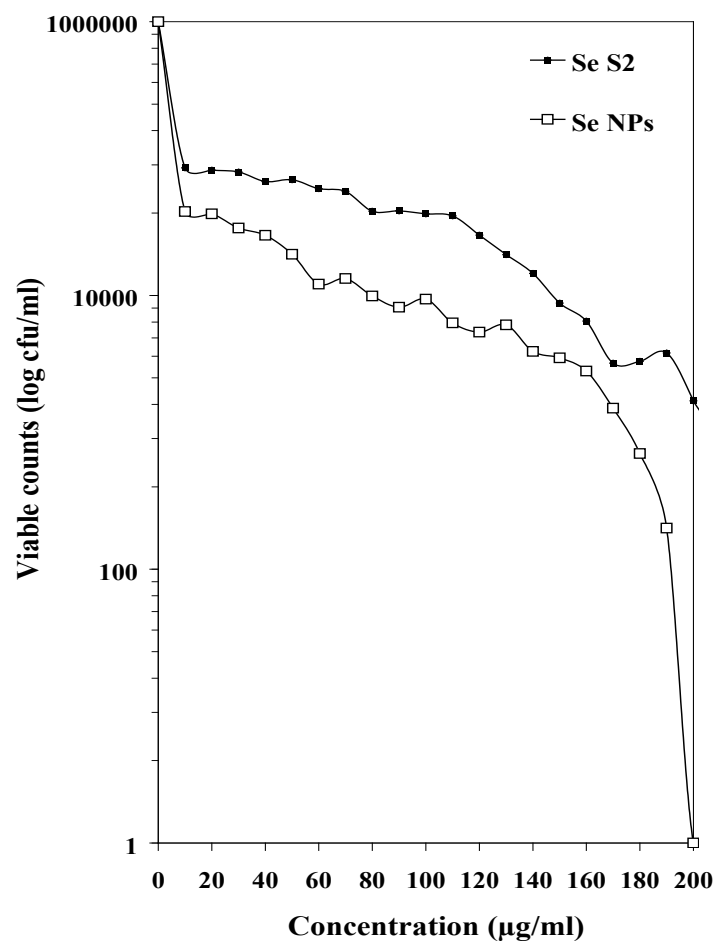


Fig1. The viability of *M. sympodialis* cells after growth with different concentrations of biogenic SeNPs and SeS₂ as determined by CFU.

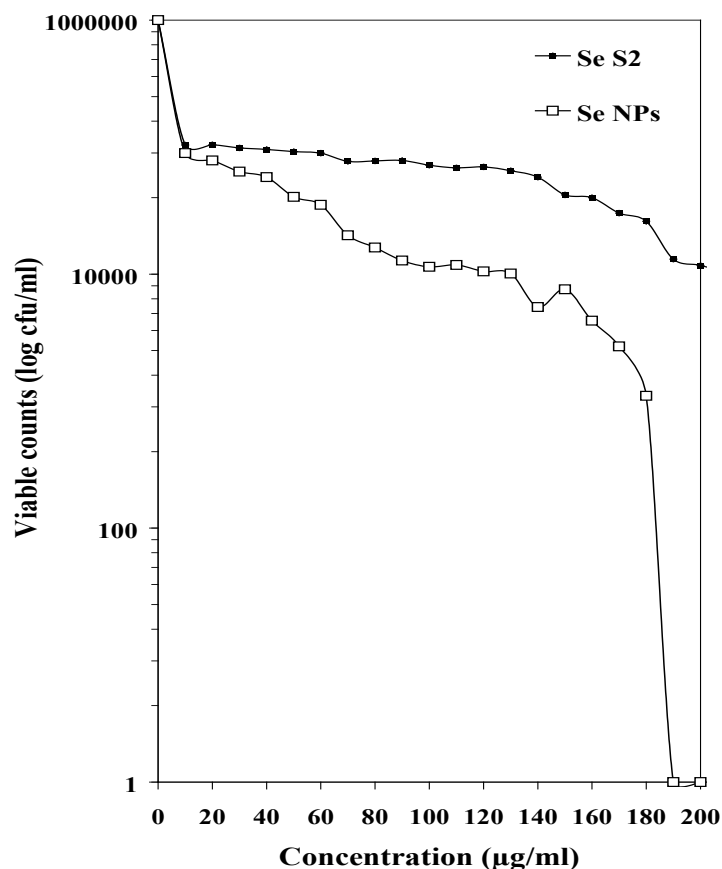


Fig2. The viability of *M. furfur* cells after growth with different concentrations of biogenic SeNPs and SeS₂ as determined by CFU.

The addition of as little as 10µg/ml of SeNPs or SeS₂ to the cultures resulted in a decrease in the number of viable cells. As the concentrations of the compounds increased from 10µg/ml to 200µg/ml, the number of surviving CFUs of both fungal species decreased proportionally, and the viable count obtained in cultures treated with each concentration of SeNPs was lower than that obtained in the cultures treated with SeS₂ (p<0.05). Whereas a low number of cells from both fungal species survived in the cultures containing 200µg/ml of SeS₂, no viable cells of these fungi were found in the cultures grown with the same concentration of SeNPs (Figure 1, Figure 2), indicating the higher antifungal potency of SeNPs.

The MICs and MFCs of SeNPs and SeS₂ were also calculated and compared (Table 1). The MIC of SeNPs was three times lower than that of SeS₂ for *M. furfur* and eight times lower than that of SeS₂ for *M. sympodialis* (Table 1). The MFCs of SeS₂ for *M. sympodialis* and *M. furfur* were 220 and 260µg/ml, respectively, and higher than the MFC of SeNPs for eliminating these fungi (Table 1). So, our data show that the fungicidal effect of SeNPs was higher than that of SeS₂. The MFCs of both compounds were slightly higher for *M. furfur* than for *M. sympodialis*.

Table1. The MIC and MFC of SeNPs and SeS₂ against *M. sympodialis* and *M. furfur*.

MFC (µg/ml)		MIC (µg/ml)		Test strain
SeS ₂	Se NPs	SeS ₂	Se NPs	
260	190	150	50	<i>M. furfur</i>
220	180	80	10	<i>M. sympodialis</i>

Antifungal Activity of Selenium Nanoparticles and Selenium Disulfide Against Two *Malassezia* Species

To investigate if exposure to SeNPs and SeS₂ resulted in the suppression of later fungal growth, the growth of *M. furfur* treated with 50 µg/ml of SeS₂ (0.33 x MIC) and SeNPs (1 x MIC) was compared with the growth of *M. furfur* in the cultures grown without selenium.

In our experiments, exposure of *M. furfur* to 50 µg/ml of either SeS₂ or SeNPs for 2 hours did not suppress growth in cultures with 5 µg/ml of either SeS₂ or SeNPs. After 120 hours of incubation, the viable count of untreated *M. furfur* grown in media without selenium was 12 x 10⁶ CFU, but for those treated cells grown with 5 µg/ml of either SeS₂ or SeNPs, the counts were 18 x 10⁶ and 19 x 10⁶ cells of *M. furfur*, respectively, indicating that neither of the preparations had a PAFE for this fungus (Figure 3).

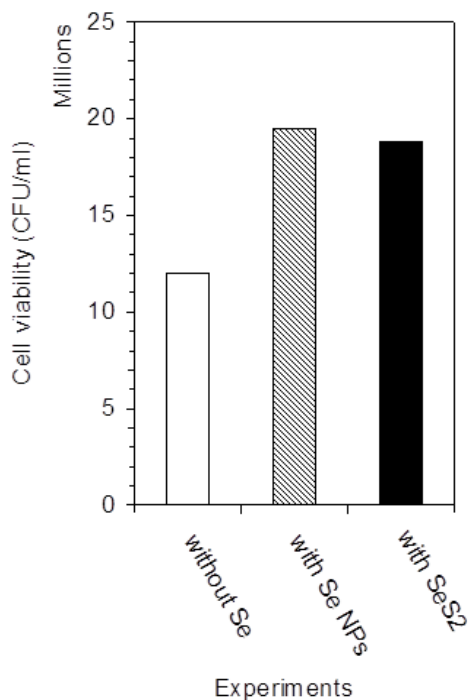


Fig3. The effect of 2 h pre-exposure of *M. furfur* to 50 µg/ml of biogenic SeNPs and SeS₂ on its regrowth with 5 µg/ml of the compound, as compared with control cultures grown in medium without selenium. All flasks were inoculated with 5 × 10⁶ cells/ml. (Data of each group are presented as mean ±SD (P = <0.001).

Drug PAFE, a suppression of fungal growth after limited drug exposure, has been observed for various compounds in some fungi [Vitale et al.,2003]. Vitale et al showed that the PAFE of amphotericin B for several fungi is dependent on concentration; the effect is not seen for all fungi. Moreover, in our experience for *M. furfur* neither of the preparations had a PAFE for this fungus (Figure 3).

Figure 2 shows that a large number of *M. furfur* cells survived after 120 h incubation following a pre-treatment for 2 hours with 50 µg/ml of either SeS₂ or SeNPs. It is possible that the original inoculum contained a population of cells with different degrees of sensitivity to selenium. The surviving cells were either resistant to 50 µg/ml of selenium originally or became resistant to selenium during incubation. In either case, they may have been metabolically different from the rest of the cells in the population. The better growth observed for cells of *M. furfur* exposed to 50 µg/ml of SeS₂ and SeNPs and then grown in the presence of 5 µg/ml of SeS₂ and SeNPs (Figure 3) could be the result of the elimination of less tolerant cells and growth of more resistant cells that could proliferate more rapidly. Development of resistance to selenium has been shown in other fungi [Buxton et al., 1989]. Selenate resistance was found in an *Aspergillus* species mutant that lacked ATP sulfurylase and was

Antifungal Activity of Selenium Nanoparticles and Selenium Disulfide Against Two *Malassezia* Species

not able to use sulfate as a sole sulfur source [Buxton et al., 1989] Selenium-resistant strains of yeast also have been found [Kitajima et al., 2013; Berdicevsky et al., 1993].

SeS₂ is commercially available as an antifungal agent in over-the-counter shampoos for the treatment of dandruff and seborrheic dermatitis. To avoid skin irritation, users are recommended to limit their exposure time to SeS₂ [Chu et al., 1984; McGinley et al., 1982; Van Cutsem et al., 1990; Hersle et al., 1971]. The proliferation of *Malassezia* spp. after short exposure to selenium in our study should be considered in regard to the treatment of fungal infection with selenium shampoos. The survival and the growth of viable *Malassezia* spp. cells after short treatment may be the cause of recurrent infection.

ACKNOWLEDGMENT

This work was supported by Deputy of Research, Tehran University of Medical Sciences, Tehran, Iran (Grant no: 89-01-33-10319). The views presented in this manuscript do not necessarily reflect those of the U.S. Food and Drug Administration.

REFERENCES

1. Aggarwal K, Jain V, Sangwan S. Comparative study of ketoconazole versus selenium sulphide shampoo in pityriasis versicolor. *IJDVL* 2003;69:86.
2. Anwar MF, Yadav D, Jain S, Kapoor S, et al. Size-and shape-dependent clinical and mycological efficacy of silver nanoparticles on dandruff. *Int J Nanomedicine* 2016;11:147.
3. Roques C, Brousse S, Panizzutti C. In vitro antifungal efficacy of ciclopirox olamine alone and associated with zinc pyrithione compared to ketoconazole against *Malassezia globosa* and *Malassezia restricta* reference strains. *Mycopathologia* 2006;162:395-400.
4. Rudramurthy SM, Honnavar P, Dogra S, Yegneswaran PP, et al. Association of *Malassezia* species with dandruff. *Indian J. Med. Res* 2014;139:431.
5. Chu, A. Comparative clinical trial of bifonazole solution versus selenium sulphide shampoo in the treatment of pityriasis versicolor. *Dermatology* **1984**, 169, 81-87.
6. McGinley KJ, Leyden JJ. Antifungal activity of dermatological shampoos. *Arch Dermatol Res* 1982;272:339-342.
7. Van Cutsem J, Van Gerven F, Franssen J, Schrooten P, et al. The in vitro antifungal activity of ketoconazole, zinc pyrithione, and selenium sulfide against *Pityrosporum* and their efficacy as a shampoo in the treatment of experimental pityrosporiasis in guinea pigs. *J Am Acad Dermatol* 1990;22:993-998.
8. Kieliszek M, Błażej S, Bzducha-Wrobel A. Influence of selenium content in the culture medium on protein profile of yeast cells *Candida utilis* ATCC 9950. *Oxid Med Cell Longev* 2015;2015:1-7.
9. Kitajima T, Chiba Y. Selenomethionine metabolism and its toxicity in yeast. *Biomol Concepts* 2013;4:611-616.
10. Wu Z-l, Yin X-b, Lin Z-q, Banuelos GS, et al. Inhibitory effect of selenium against *Penicillium expansum* and its possible mechanisms of action. *Curr Microbiol* 2014;69:192-201.
11. Oremland RS, Herbel MJ, Blum JS, Langley S, et al. Structural and spectral features of selenium nanospheres produced by Se-respiring bacteria. *Appl Environ Microbiol* 2004;70:52-60.
12. Ren G, Hu D, Cheng EW, Vargas-Reus MA, et al. Characterisation of copper oxide nanoparticles for antimicrobial applications. *Int J Antimicrob Agents* 2009;33:587-590.

Antifungal Activity of Selenium Nanoparticles and Selenium Disulfide Against Two *Malassezia* Species

13. Sadiq IM, Chowdhury B, Chandrasekaran N, Mukherjee A. Antimicrobial sensitivity of *Escherichia coli* to alumina nanoparticles. *Nanomedicine: NBM* 2009;5:282-286.
14. Tan L, Jiang X, Zhang Y, Tang H, et al. In vitro study on the individual and synergistic cytotoxicity of adriamycin and selenium nanoparticles against Bel7402 cells with a quartz crystal microbalance. *Biosens Bioelectron* 2009;24:2268-2272.
15. Wang H, Wei W, Zhang SY, Shen YX, et al. Melatonin-selenium nanoparticles inhibit oxidative stress and protect against hepatic injury induced by *Bacillus Calmette-Guerin*/lipopolysaccharide in mice. *J Pineal Res* 2005;39:156-163.
16. Wang H, Zhang J, Yu H. Elemental selenium at nano size possesses lower toxicity without compromising the fundamental effect on selenoenzymes: comparison with selenomethionine in mice. *Free Radic Biol Med* 2007;42:1524-1533.
17. Gajjar P, Pettee B, Britt DW, Huang W, et al. Antimicrobial activities of commercial nanoparticles against an environmental soil microbe, *Pseudomonas putida* KT2440. *J Biol Eng* 2009;3:9.
18. Zeng H, Combs GF. Selenium as an anticancer nutrient: roles in cell proliferation and tumor cell invasion. *J. Nutr. Biochem* 2008;19:1-7.
19. Zhang J-S, Gao X-Y, Zhang L-D, Bao Y-P. Biological effects of a nano red elemental selenium. *Biofactors* 2001;15:27-38.
20. Shahverdi A, Fakhimi A, Mosavat G, Jafari-Fesharaki P, et al. Antifungal activity of biogenic selenium nanoparticles. *WASJ* 2010;10:918-922.
21. Fesharaki PJ, Nazari P, Shakibaie M, Rezaie S, et al. Biosynthesis of selenium nanoparticles using *Klebsiella pneumoniae* and their recovery by a simple sterilization process. *Braz J Microbiol.* 2010;41:461-466.
22. Espinel-Ingroff A, Fothergill A, Peter J, Rinaldi M, et al. Testing conditions for determination of minimum fungicidal concentrations of new and established antifungal agents for *Aspergillus* spp.: NCCLS collaborative study. *J Clin Microbiol* 2002;40:3204-3208.
23. Vitale RG, Meis JF, Mouton JW, Verweij PE. Evaluation of the post-antifungal effect (PAFE) of amphotericin B and nystatin against 30 zygomycetes using two different media. *J Antimicrob Chemother* 2003;52:65-70.
24. Buxton FP, Gwynne DI, Davies RW. Cloning of a new bidirectionally selectable marker for *Aspergillus* strains. *Gene* 1989;84:329-334.
25. Berdicevsky I, Duek L, Merzbach D, Yannai S. Susceptibility of different yeast species to environmental toxic metals. *Environ Pollut* 1993;80:41-44.
26. Hersle K. Selenium sulphide treatment of tinea versicolor. *Acta Derm Venereol* 1971;51:476-478.

Citation: Faranak Mavandadnejad, Fatemeh Rafii, Elnaz Faghfuri, Narges Mokhtari-Nori, Sasan Rezaie, Ahmad Reza Shahverdi, "Antifungal Activity of Selenium Nanoparticles and Selenium Disulfide Against Two *Malassezia* Species". *American Research Journal of Dermatology*. 1(1): 22-28.

Copyright © Faranak Mavandadnejad, Fatemeh Rafii, Elnaz Faghfuri, Narges Mokhtari-Nori, Sasan Rezaie, Ahmad Reza Shahverdi, This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.